Assessment of serum dehydroepiandrosterone sulphate in subjects during the pre-pubertal, pubertal, and adult stages of skeletal maturation

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SUMMARY The aim of this cross-sectional study was to evaluate serum levels of the hormone, dehydroepiandrosterone sulphate (DHEAS), during the pre-pubertal, pubertal, and adult stages of skeletal maturation based on the methods of Björk and Grave and Brown of assessing hand—wrist radiographs. The levels of the DHEAS of each individual were measured using quantitative enzyme-linked immunosorbent assay and correlated with the corresponding stages in their hand—wrist radiograph. This study was performed on 60 subjects (30 females and 30 males) aged from 7 to 30 years.

Analysis of variance followed by a Tukey honestly significant difference test showed that the serum levels of the DHEAS were statistically significant at (P < 0.01) in all three groups. The serum levels were significant (P < 0.05) when each of the three groups were individually compared with the other two groups. The mean DHEAS levels were 0.43 ± 0.28 , 2.17 ± 0.92 , and $4.60 \pm 1.34 \,\mu\text{g/ml}$ in the pre-pubertal, pubertal, and adult groups, respectively. There was a gradual increase in the hormonal level with progressing skeletal age. The adult group showed the highest DHEAS level and the pre-pubertal group the lowest.

Serum levels of DHEAS showed a constant increase from pre-puberty to adulthood, and at the same level of skeletal maturation, both females and males had similar hormone levels. This finding highlights the fact that the hormone DHEAS plays a significant role and can be a valuable tool in assessing skeletal maturation.

Introduction

Appropriate timing of the interception of a skeletal malocclusion is a key to success in dentofacial orthopaedics. Growth assessment is carried out using various skeletal maturity assessment tools such as hand-wrist and cervical vertebrae radiographs (Bambha and Natta, 1963; Björk and Helm, 1967; Mitani, 1977; Moore et al., 1990). Skeletal maturation accelerates during puberty. There is a strong correlation between craniofacial growth and somatic changes in puberty (skeletal maturation; Nanda, 1955). Puberty is primarily a neuroendocrinal event, with the pituitary and hypothalamus playing major roles in its initiation. Both the pituitary and hypothalamus are together called 'gonadostat'. Puberty is marked by the pulsatile secretion of the gonadotropin-releasing hormone from the hypothalamus, which stimulates the pituitary to secrete gonadotropins (follicle stimulating and luteinizing hormones). Gonadotropins stimulate the gonads to secrete the sex steroids testosterone and oestrogen, which in turn affects somatic changes or skeletal maturation in puberty (Carlson and Ribbens, 1986).

For the gonadostat to initiate its action, it requires stimulation from the adrenal gland, which secretes a significant amount of a steroid, dehydroepiandrosterone (DHEA), and its sulphated conjugate dehydroepiandrosterone sulphate (DHEAS; Auchus and Rainey 2004). They are present in circulation approximately 3 years prior to puberty. This period is termed as adrenarche (Sizonenko and Paunier 1975; Carlson and Ribbens 1986; Remer *et al.*, 2004) and the steroids DHEA and DHEAS are responsible for the stimulation of the gonadostat to take effect. Both DHEA and DHEAS have similar actions. They are found to stimulate growth and proliferation of epiphyseal cartilage and potentiate the action of growth hormone (GH). The phenotypic result of adrenarche is pubarche or the development of secondary sexual characteristics such as axillary and pubic hair that occurs in both girls and boys at approximately 11 years of age. Premature and exaggerated adrenarche can be indicative of future onset of adult diseases, thus increasing clinical relevance.

DHEAS has also been found to enhance bone deposition (Christian *et al.*, 1997; Remer *et al.*, 2003; Bonofiglio *et al.*, 2004; Auchus and Rainey, 2004; Adachi and Takayanagi, 2006), to increase bone mineral density, and to maintain the cancellous and cortical bone mass by way of protective action in osteoblasts (Bing *et al.*, 1988; Bonofiglio *et al.*, 2004; Remer *et al.*, 2004; Adachi and Takayanagi, 2006). Christian *et al.* (1997) stated that DHEA and DHEAS have actions similar to dehydrotestosterone on human osteoblast

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cell metabolism. They exert their mitogenic influence on osteoblasts through androgen receptor-mediated mechanisms and stimulate the action of alkaline phosphatase activity through tumour growth factor β expression.

DHEAS is the immediate metabolite of DHEA and the concentration of DHEAS is 100- to 1000-fold greater than DHEA. Unlike DHEA, DHEAS does not show any diurnal variation. Hence, measurement of the serum levels of DHEAS is more reliable and convenient when compared with measurement of DHEA.

As hormones play a primary role in the initiation of puberty, it would be more appropriate to measure their serum levels (the cause) rather than study the effects of hormones or puberty. This study aimed to determine the relationship between serum levels of DHEAS and skeletal maturation assessed using hand—wrist radiographs.

Subjects and methods

Sixty individuals (30 males, 30 females) between 7 and 30 years of age (mean 22.95 years) participated in the cross-sectional study. All individuals had a Class I occlusion. Subjects with any systemic disease or under any medication were excluded from the study. Informed consent was obtained from the selected individuals and/or their parents. The study was approved by the institutional ethical committee of Madras Medical College, Chennai.

The individuals were divided into three groups: 20 prepubertal (Figure 1A), pubertal (Figure 1B), and adult (Figure 1C) based on the skeletal maturation stage assessed using the method of Björk (1972) and Grave and Brown (1976). Each group contained 10 males and 10 females. Hand—wrist radiographs were taken with 60 mA X-ray machine with 40 kV and 12–16 mAs speed using extra-oral radiographic film (Kodak T-mat Blue; Kodak Limited, Hemel Hempstead, Hertfordshire, UK). The radiographs were taken by placing the cassette with its long axis parallel to the long axis of the hand. The subjects were seated on an adjustable stool with their left forearm resting on the table and the palm of the hand facing downward with the lower

end of the radius and ulna also included on the film. The radiographs were evaluated in a dark room on a cephalometric table with posterior illumination and traced on an acetate paper by the same author (BS). Evaluation of hand—wrist radiographs was carried out using the method of Björk (1972) and Grave and Brown (1976).

Estimation of the concentration of serum DHEAS was undertaken using quantitative enzyme-linked immunoassay (ELISA kit, Dia Metra, Milano, CE Italy). Approximately 2.5 ml of venous blood was collected from each patient and centrifuged for serum separation. The samples were stored in Ependorf tubes at -20°C.

The standard curve (Figure 2) for hormone concentration was plotted in the X-axis and absorbance on the Y-axis using the set of six standards provided in the ELISA kit, by a semi-automated ELISA reader. When the absorbance values of the samples had been obtained, the corresponding DHEAS concentrations were calculated from the standard curve on the X-axis. The measured serum DHEAS values for each group were calculated. Analysis of variance was used to compare the mean hormone value of the three groups followed by a Tukey honestly significant difference (HSD) test to assess the level of significance for multilevel comparison. A Student's *t*-test was used to compare the difference between the males and females in each group.

Results

The distribution of the hormone levels in the pre-pubertal, pubertal, and adult groups is shown in Figures 3A, 3B, and 3C. The mean DHEAS values in each group were 0.43 (pre-pubertal), 2.17 (pubertal), and 4.60 μ g/ml (adult). The standard deviation for each of the groups was 0.28, 0.92, and 1.34, respectively. There was a significant difference (P < 0.001) in the means of hormone levels between the groups (Table 1).

Tukey HSD test showed that each group was significantly different from the other two groups (Table 1). The 95 per cent confident interval for mean for the hormone levels in







Figure 1 Hand-wrist radiograph of a pre-pubertal (A), pubertal (B), and adult (C) subject based on the methods of assessment of Björk and Grave and Brown.

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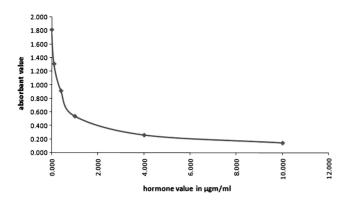
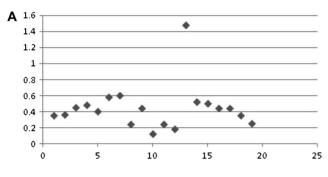
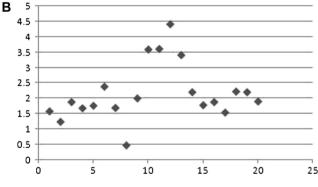


Figure 2 Standard curve obtained for the hormone, dehydroepiandrosterone sulphate, using enzyme-linked immunosorbent assay. The hormone levels are marked on *X*-axis and the absorbant values in Y-axis.





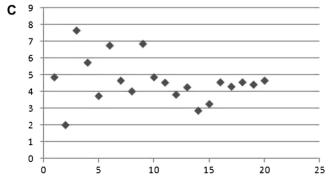


Figure 3 Scatter diagrams depicting the distribution of dehydroepiandrosterone sulphate values in (A) the pre-pubertal, (B) pubertal, and (C) adult groups.

each group were pre-pubertal: 0.2993–0.5627 $\mu g/ml$; pubertal: 1.7388–2.60002 $\mu g/ml$ and adult: 3.9732–5.2268 $\mu g/ml$.

Table 1 Analysis of variance to test the significance in serum dehydroepiandrosterone sulphate among the pre-pubertal, pubertal, and adult groups.

| Groups | Hormone value | | P value |
|--------------|---------------|------|---------|
| | Mean | SD | |
| Pre-pubertal | 0.43* | 0.28 | |
| Pubertal | 2.17* | 0.92 | ** |
| Adult | 4.60* | 1.34 | |

^{*}Tukey honestly significant difference indicates significant at the 5 per cent level when an individual group was compared with the other two groups.

Table 2 Comparison of the hormone dehydroepiandrosterone sulphate levels between male and female subjects in the prepubertal, pubertal, and adult groups.

| Groups | Gender | Mean ± SD | P value |
|--------------|--------|-----------------|---------|
| Pre-pubertal | Male | 0.48 ± 0.38 | 0.489 |
| | Female | 0.39 ± 0.14 | |
| Pubertal | Male | 2.17 ± 1.21 | 0.998 |
| | Female | 2.17 ± 0.56 | |
| Adult | Male | 4.93 ± 1.81 | 0.276 |
| | Female | 4.27 ± 0.53 | |

The mean DHEAS was pre-pubertal group: male $0.48\pm0.38\,\mu\text{g/ml}$; female $0.39\pm0.14\,\mu\text{g/ml}$, pubertal group: male $2.17\pm1.21\,\mu\text{g/ml}$; female $2.17\pm0.56\,\mu\text{g/ml}$, and adult group: male $4.93\pm1.81\,\mu\text{g/ml}$; female $4.27\pm0.53\,\mu\text{g/ml}$. There was no significant difference in hormone levels between males and females in any group at a particular stage of skeletal maturation at the 5 per cent level (Table 2).

Discussion

Time, the fourth dimension in orthodontics, is important when planning growth modification therapy. Successful growth modification is related to the amount of remaining growth and accurate prediction of growth spurts. The adolescent growth spurt is the most important for orthodontic treatment because the physical changes at adolescence significantly affect the face and dentition (Nanda, 1955; Hunter, 1966).

Earlier methods of assessing growth included physical stature, peak height velocity, and growth charts. All these methods assessed growth in relation to chronological age. These were later followed by using skeletal maturation as an indicator of physical development and maturation, which led to the radiographic method of determining skeletal maturation (Ranke, 1896; Todd, 1930). Greulich and Pyle (1959) compiled a radiographic atlas of skeletal development of the hand and wrist, which were developed on the basis of skeletal age as opposed to chronological age (Carlson and Ribbens, 1986).

^{**}P < 0.001

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Timing of tooth eruption was related to skeletal maturation by Engstrom *et al.* (1983) who reported a correlation between lower third molar development, skeletal maturation, and chronological age. Nanda (1960), Lewis (1991), and Demirjian *et al.* (1985) found less association between the time of tooth eruption and skeletal maturation. Conversely, Coutinho *et al.* (1993) stated that canine calcification can be used as a tool for assessing skeletal maturation, in agreement with the findings of Chertkow and Fatti (1979).

Hellman (1927), Björk and Helm (1967), Houston *et al.* (1979), Singer (1980), Fishman (1982), Hägg and Taranger (1982), Leite and O'Rielly (1987), Pileski *et al.* (1973), and Abdel-Khader (1998) associated the ossification events of the hand—wrist bones with the pubertal growth spurt. Although other bones such as the carpals, the femur, the elbow joint, the shoulder joint, and the skull can be used to assess skeletal maturation, the hand—wrist proved to be a more effective method because of numerous centres of ossification which undergo changes at different times and rates.

Cervical vertebrae are also used for assessment of skeletal maturation (Lamparski, 1975; Hassel and Farman, 1995; Fernandez *et al.*, 1998; Franchi *et al.*, 2000; Baccetti *et al.*, 2000). The advantages of the cervical vertebrae include no additional radiation as they can be visualized on a lateral cephalometric radiograph. The disadvantages include difficulties in visualization of the subtle changes in the vertebrae, in visualization due to incorrect neck posture while taking the radiograph, and blocking out of cervical vertebrae due to the use of a thyroid collar (Leite and O'Rielly, 1987). The frontal sinus has also been used as an indicator of the pubertal growth spurt (Rossouw *et al.*, 1991; Ruf and Pancherz, 1996).

Puberty is under the control of the neuroendocrine system. Pubertal maturation is initiated by the central nervous system with increased secretion of gonadotropin-releasing hormone, gonadotropins (follicle-stimulating hormone and luteinizing hormones), sex steroids, GH, and somatomedin C. The neuroendocrine role in the onset of puberty starts with maturation of the hypothalamus pituitary complex, which is heralded by a significant increase in the secretion of DHEA and its conjugate DHEAS (Auchus and Rainey 2004). Serum levels of DHEAS are high in the neonate, after which there is a decrease, then a rapid increase in the serum level from 7 years of age in females and 8 years of age in males, with a gradual increase until it attains the adult value (Hopper and Yen, 1975; Sulcova et al., 1984). The first peak in DHEAS concentration occurs between 6 and 8 years of age in both males and females and the second peak occurs at 11 years in females and 13 years in males (Sizonenko and Paunier, 1975; Rotteveel et al., 1997; Kulik-Rechberger et al., 2000).

Only one study has been conducted on the levels of DHEAS with regard to monitoring growth during orthodontic treatment (Ghafari *et al.*, 1995). The results of the present study clearly show that serum concentration of DHEAS for the pre-pubertal, pubertal, and adult groups was statistically significant. There

was a progressive rise in serum concentration as skeletal maturation progressed, almost reaching a maximum value after the complete fusion of the epiphysis and diaphysis of the radius. The results also showed that low levels of DHEAS were present in the pre-pubertal group with increased values in the pubertal group and the highest levels in the adult group, showing a gradual increase as maturation progresses. Within each group, there was no significant difference in the mean hormone values between males and females, demonstrating that there was no gender difference in the hormone values at a particular stage of skeletal maturation. This finding is in contrast to the results of Ghafari et al. (1995) who found a significant gender difference in DHEAS level of the same age. The difference in DHEAS levels between males and females apparently reflects gender differences related to the earlier maturation of girls than boys. The difference in the findings could be due to the fact that in the present study, DHEAS levels were compared based on skeletal maturation and not on chronological age. Another finding of note was the individual variation in the DHEAS levels within the groups. In addition, some individuals in the pubertal group showed higher levels of DHEAS than those in the adult group. This could be attributed to a higher body mass index (l'Allemand et al., 2002), which was not measured in the present study.

The evaluation of hormones can be repeated without the disadvantages of radiation exposure and can be charted to assess and predict skeletal maturation. The disadvantage of this procedure is the invasive venipuncture. However, non-invasive procedures such as evaluation of salivary DHEAS has proved to be unreliable (Read, 1993).

DHEA and DHEAS are considered as the markers of adrenarche. Their influence on bone growth and correlation with the stages of hand—wrist radiographs make them useful as a possible indicator of skeletal maturation to aid the assessment of growth status during adolescence.

Conclusions

The present study showed that serum levels of DHEAS in the pre-pubertal, pubertal, and adult groups were statistically significant. The findings also confirm that DHEAS is associated with growth during the pubertal growth spurt and possibly plays a direct role in skeletal maturation.

Acknowledgements

The authors acknowledge the help of Dr K S G A Nasser, and Professors W S Manjula and M C Sainath, from Tamil Nadu Government Dental College and Hospital, Chennai in completing the study.

References

Abdel-Khader H M 1998 The reliability of dental X-ray film in assessment of MP3 stages of pubertal growth spurt. American Journal of Orthodontics and Dentofacial Orthopedics 114: 427–429 DHEAS AND SKELETAL MATURATION 451

- Adachi M, Takayanagi R 2006 Role of androgens and DHEA in bone metabolism. Clinical Calcium 16: 61–66
- Auchus R J, Rainey W E 2004 Adrenarche—physiology, biochemistry, and human disease. Journal of Clinical Endocrinology 60: 288–296
- Baccetti T, Franchi Z, Toth Z R 2000 Treatment timing of twin block therapy. American Journal of Orthodontics and Dentofacial Orthopedics 118: 159–170
- Bambha J K, Natta P V 1963 A longitudinal study of occlusion and tooth eruption in relation to skeletal maturation. American Journal of Orthodontics 49: 481–493
- Bing C, Xu S E, Zhang G D, Wang W Y 1988 Serum dehydroepiandrosterone sulphate and pubertal development in Chinese girls. Annals of Human Biology 15: 421–429
- Björk A 1972 Timing of interceptive orthodontic measures based on stages of maturation. Transactions of the European Orthodontic Society 61–74
- Björk A, Helm S 1967 Prediction of age of maximum pubertal growth in body height. Angle Orthodontist 37: 134–143
- Bonofiglio D, Garofalo C, Catalano S, Marisco S, Aquila S, Ando S 2004 Low calcium intake is associated with decreased adrenal androgens and reduced bone age in premenarcheal girls in the last pubertal stages. Journal of Bone and Mineral Metabolism 22: 64–70
- Carlson D S, Ribbens K A 1986 Craniofacial growth during adolescence, Monograph No 20, Craniofacial Growth Series, Center for Human Growth and Development. University of Michigan, Ann Arbor, pp. 1–207
- Chertkow S, Fatti P 1979 The relationship between tooth mineralization and early radiographic evidence of the ulnar sesamoid. Angle Orthodontist 49: 282–288
- Christian H K, Glenn K W, Hierl T, Zeigler R 1997 Gonadal and adrenal androgens are potent regulators of human bone cell metabolism *in vitro*. Journal of Bone and Mineral Research 12: 464–471
- Coutinho S, Buschang P H, Miranda I 1993 Relationship between mandibular canine calcification stages and skeletal maturity. American Journal of Orthodontics and Dentofacial Orthopedics 104: 262–268
- Demirjian A, Buschang P H, Tanguay R, Pattersen D K 1985 Interrelationship among measure of somatic, skeletal dental and sexual maturity. American Journal of Orthodontics 88: 433–438
- Engstrom C, Engstrom H, Sagne S 1983 Lower third molar development in relation to skeletal maturity and chronological age. Angle Orthodontist 53: 97–106
- Fernandez G P, Torre H, Flores L, Rea J 1998 The cervical vertebrae as maturational indicators. Journal of Clinical Orthodontics 32: 221–225
- Fishman L S 1982 Radiographic evaluation of skeletal maturation. A clinically oriented method based on hand-wrist films. Angle Orthodontist 52: 88–112
- Franchi L, Baccetti T, McNamara J A Jr. 2000 Mandibular growth as related to cervical vertebral maturation and body height. American Journal of Orthodontics and Dentofacial Orthopedics 118: 335–340
- Ghafari J G, Shofer F S, Laster L L, Markowitz D L, Silverton S, Katz S H 1995 Monitoring growth during orthodontic treatment. Seminars in Orthodontics 1: 165–175
- Grave K C, Brown T 1976 Skeletal ossification and adolescent growth spurt. American Journal of Orthodontics 69: 611-619
- Greulich W W, Pyle S I 1959 Radiographic atlas of skeletal development of the hand and wrist, 2nd edn.. Stanford University Press, Stanford, pp. 1–256
- Hägg U, Taranger J 1982 Maturation indicators and pubertal growth spurt. American Journal of Orthodontics 82: 299–309
- Hassel B, Farman A G 1995 Skeletal maturation evaluation using cervical vertebrae. American Journal Orthodontics and Dentofacial Orthopedics 107: 58–66
- Hellman M 1927 The face and occlusion of teeth in man. International Journal of Orthodontics and Orthognathic Surgery 13: 921–945
- Hopper B R, Yen S S C 1975 Circulating concentrations of dehydroepiandrosterone and dehydroepiandrosterone sulfate during puberty. Journal of Clinical Endocrinology and Metabolism 40: 458–461

Houston W J B, Miller J C, Tanner J M 1979 Prediction of the timing of the adolescent growth spurt from ossification events in hand wrist films. British Journal of Orthodontics 6: 145–152

- Hunter C J 1966 The correlation of facial growth with both body height and skeletal maturation at adolescence. Angle Orthodontist 36: 44–54
- Kulik-Rechberger B, Furmaga-Jablonska W, Rechberger T 2000 The role of dehydroepiandrosterone sulphate during puberty in girls. Ginekelgia Polska 71: 668–672
- l'Allemand D, Schimdt S, Rousson V, Brabant G, Gasser T, Gruters A 2002 Associations between body mass, leptin IGF-1, and circulating adrenal androgens in children with obesity and premature adrenarche. European Journal of Endocrinology 146: 537–543
- Lamparski D G 1975 Skeletal age assessment utilizing cervical vertebrae. American Journal of Orthodontics 67: 458–459
- Leite H R, O'Rielly M I 1987 Skeletal age assessment using the 1st, 2nd, 3rd finger of the hand. American Journal of Orthodontics and Dentofacial Orthopedics 92: 492–498
- Lewis A B 1991 Comparison between dental and skeletal ages. Angle Orthodontist 61: 87–92
- Mitani H 1977 Occlusal and craniofacial growth changes during puberty. American Journal of Orthodontics 72: 77–84
- Moore R N, Moyer B A, Du Bois L M 1990 Skeletal maturation and craniofacial growth. American Journal of Orthodontics and Dentofacial Orthopedics 98: 33–40
- Nanda R S 1955 The ratio of the growth of several facial components measured from serial cephalometric roentgenogram. American Journal of Orthodontics 41: 658–673
- Nanda R S 1960 Eruption of human teeth. American Journal of Orthodontics 46: 363–378
- Pileski R C A, Woodside D G, James G A 1973 Relationship of ulnar sesamoid bone and maximum mandibular growth velocity. Angle Orthodontist 43: 162–170
- Ranke J 1896 Uber Die Ossifikation. Munchen Medizinische Wochenschrift 43: 686
- Read G F 1993 Status report on measurement of salivary estrogens and androgens. Annals of the New York Academy of Sciences 694: 146–160.
- Remer T et al. 2003 Adrenarche and bone modeling and remodeling at the proximal radius: weak androgens make stronger cortical bone in healthy children. Journal of Bone and Mineral Research 18: 1539–1546
- Remer T et al. 2004 Adrenal steroid hormones and metaphyseal bone in children. Journal of Hormonal Research 62: 221–226
- Rossouw P E, Lombard C J, Harris A M P 1991 The frontal sinus and mandibular growth prediction. American Journal of Orthodontics and Dentofacial Orthopedics 100: 542–554
- Rotteveel J, de Ridder C, Schoute E, Delemarre-van de Waal H A 1997 Androstenedione, dehydroepiandrosterone sulphate and estradiol levels throughout puberty: relation to peak height velocity. Journal of Hormone Research 48: 263–267
- Ruf S, Pancherz H 1996 Frontal sinus development as indicator for somatic maturity at puberty. American Journal of Orthodontics and Dentofacial Orthopedics 110: 476–482
- Singer J 1980 Physiological timing of orthodontic treatment. Angle Orthodontist 50: 323–333
- Sizonenko P C, Paunier L 1975 Hormonal changes in puberty. III: correlations of plasma dehydroepiandrosterone, testosterone, FSH, and LH with stages of puberty and bone age in normal boys and girls and in patients with Addison's disease or hypogonadism or with premature or late adrenarche. Journal of Clinical Endocrinology and Metabolism 41: 894–904
- Sulcova J, Hill M, Hampl R, Starka L 1984 Age and sex related differences in serum levels of unconjugated dehydroepiandrosterone and its sulphate in normal subjects. Journal of Endocrinology 154: 57–62
- Todd T W 1930 The roentgenographic appraisement of skeletal differentiation. Child Development 1: 298–310